

# Multi-Mycotoxin Contaminations in Fish Feeds from Different Agro-Ecological Zones in Nigeria

Momodu Foluke Olorunfemi<sup>1</sup>, Adegboyega Christopher Odebode<sup>1</sup>, Olawuyi Odunayo Joseph<sup>2</sup>, Chibundu Ezekiel<sup>2</sup>, Michael Sulyok<sup>3</sup>, Rudolf Krska<sup>3</sup>, Adedayo Oyedele<sup>4</sup>

<sup>1</sup>University of Ibadan, Dept. of Botany, Nigeria, <sup>2</sup>Babcock University, Biosciences and Biotechnology, Nigeria, <sup>3</sup>University of Natural Resources and Life Sciences, Dept. of Agrobiotechnology, Austria, <sup>4</sup>Institute of Agricultural Research and Training (IAR&T), Land and Water Resources Management Programme, Nigeria

Corresponding Author: [graceplusplus@yahoo.com](mailto:graceplusplus@yahoo.com)

## INTRODUCTION

Fish and fish products constitute about 49% of the animal protein consumed in Nigeria with a population of 160,000,000 people (Ajani, 2008). There is high demand for fish as they supply a good nutritional balance of protein, vitamins and minerals with low fat of 10% calories and an excellent source of omega 3 fatty acids. The total fish demand in Nigeria stands at 2.66 million metric tonnes out of which only 0.62 million metric tonnes was produced locally (FDF, 2008), also there is dwindling amount of fish realized from fish capture as a result of overfishing and pollution (Delgado *et al.*, 2003). The Aquaculture practices in Nigeria have witnessed tremendous growth in terms of production with a record of a hundred percent increase between 2004 and 2007 (Falaye, 2013). The rise in production activities brought about the need to satisfy the high demand for the provision of household fish consumed in Nigeria. A major challenge faced in the fish farming industry is low productivity and death caused by moulds and mycotoxins contamination of fish feeds. Fish feed is one of the main input in fish farming produced by the combination of plants and animal based sources, vitamins premixes and mineral sources in right proportions to meet nutritional requirements of fishes. Fish feeds get contaminated with moulds and mycotoxins via these ingredients of plant based sources of which maize, groundnut cake, soybean cake, cassava are included, but are highly susceptible to moulds attack and consequently mycotoxins contamination. There is also the risk of introducing moulds and mycotoxins into the feeds through unhygienic feed mill environment where the ingredients are stored before formulation and during production processes (Banu and Muthumary, 2008). Moulds and mycotoxins contamination reduce feed quality and predispose fishes to incidences of mycotoxicoses when consumed. Fishes are sensitive to mycotoxins causing feeding inefficiencies and poor growth. Pelleted fish feeds when not consumed get soaked and sinks to the bottom thereby polluting the water, causing untimely deaths in fishes and resulting to financial losses to fishers, processors and sellers. This aggravates food insecurity in Nigeria. Therefore, local production of high quality fish feed is very crucial to the development and sustainability of aquaculture in Africa especially, in the rural areas. The occurrence of moulds and mycotoxins in fish feeds must be minimized to improve quality. Records of co-occurrence of mycotoxin contaminations of fish feeds are documented in Egypt (Abdelhamid *et al.*, 1998), US (Lumlertdacha and Lovell, 1995), Indonesia (Ali *et al.*, 1998), Central Europe and Switzerland (Pietsch *et al.*, 2013). Considering limited of information on toxigenic fungi from fish feeds and multimycotoxin contaminations from African countries, particularly in Nigeria, this work will make a contribution to global ecological database of toxigenic moulds and consequent multimycotoxin production from fish feeds in different Agro-Ecological Zones of Nigeria.

## MATERIALS AND METHODS

**Sampling :** Ninety four samples of fish feeds were randomly collected at different locations in the Agro Ecological Zones of Nigeria. Sampling of fish feeds from bags in warehouses and feed mills were done according to the method described by Karthik *et al.* (2009). Samples were collected at the top layer, middle and bottom layers. The bulk samples were collected into transparent Zip-lock bags and transported to the Microbiology Laboratory of Nigerian Stored Products Research Institute Ibadan, Nigeria. The samples were comminuted immediately in order to reduce particle size and stored at -20°C prior to transport to Austria for multi-mycotoxin analysis using LC-MS/MS.

## Quantification of multi-mycotoxins in fish feed samples

This was carried out by the method described by Sulyok *et al.*(2007). The analysis was done at Center for Analytical Chemistry, Department of Agrobiotechnology (IFA-Tulln), University of Natural Resources and Life Sciences Vienna, Konrad Lorenz Str, 20, A-3430 Tulln, Austria. The spectrum of mycotoxins were determined in 94 fish feed samples by LC–MS/MS. Five gram of each homogenized sample was weighed into a 50 ml polypropylene tube (Sarstedt, Germany) and extracted with 20 ml acetonitrile/water/acetic acid (79:20:1, v/v/v) for 90 min on a GFL 3017 rotary shaker (GFL, Burgwedel, Germany). The extracts were diluted and injected into the instrument as described by Sulyok *et al.* (2007). Screening of the analytes was performed using a QTrap 5500 LC–MS/MS System (Applied Biosystems, CA, US) equipped with a TurboIonSpray electrospray ionization (ESI) source and a 1290 Series HPLC System (Agilent, Waldbronn, Germany). Chromatographic separation was performed at 25 °C on a Gemini® C18-column, 150×4.6 mm i.d., 5 µm particle size, equipped with a C18 4×3 mm i.d. security guard cartridge (Phenomenex, CA, US). The identification of positive analyte was confirmed when two MRMs which yielded 4.0 identification points were obtained according to commission decision 2002/657/EC. In order to distinguish between incomplete extraction and matrix induced suppression of the LC–MS/MS signal, the apparent recovery and the recovery of the extraction step were determined by spiking three individual samples of fish feeds before extraction as well as after extraction on the same concentration level (Sulyok *et al.*, 2006).

## Statistical analysis

Data was analysed using the t-test at 95% confidence level. The mean and t-values, standard deviation and standard error of the concentrations of the multi-mycotoxins in fish feed samples were evaluated.

## RESULTS AND DISCUSSION

**Table 1: The mean and range level of mycotoxins contamination of fish feed industries from different Agro-Ecological Zones (AEZs) in Nigeria.**

Mycotoxins	GS (µg/kg)		HF (µg/kg)		DS (µg/kg)		SZ (µg/kg)		MSZ (µg/kg)		Mean Ave
	mean	range	mean	range	mean	range	mean	range	mean	range	
Aflatoxin B1	119.8	1.1-408.7	90.8	<LOD-418.7	107.4	129.6-150.8	90.4	2.4-319.0	106.8	21.8-255.5	103.0
Aflatoxin B2	14.9	2.8-53.0	11.7	<LOD-69.3	13.8	9.8-35.6	13.8	<LOD-49.0	16.6	2.4-3.0	14.2
Aflatoxin G1	46.0	<LOD-195.8	36.7	<LOD-195.8	41.9	32.9-56.2	34.0	1.3-128.3	38.0	8.7-76.1	39.3
Aflatoxin G2	4.2	<LOD-11.8	5.5	<LOD-11.8	5.3	<LOD-2.4	4.9	<LOD-4.2	4.4	<LOD-2.3	4.9
Aflatoxin M1	5.0	<LOD-17.7	4.2	<LOD-17.7	4.6	3.4-12.1	4.2	1.8-14.9	4.9	1.2-9.0	4.6
Fumonsin B1	1525.4	<LOD-6097.9	723.6	<LOD-4548.3	807.3	794.4-1035.0	787.9	64.9-755.2	660.2	<LOD-1454.5	900.9
Fumonsin B2	377.6	<LOD-1203.3	185.4	<LOD-886.0	202.8	137.2-290.6	180.0	19.7-151.4	157.0	<LOD-281.0	220.6
Fumonsin B3	66.5	<LOD-214.7	56.3	<LOD-210.9	57.6	22.4-38.6	50.7	<LOD-39.7	39.8	<LOD-48.1	54.2
Hydrolysed FB1	11.9	<LOD-53.4	8.1	<LOD-23.0	8.2	6.3-11.2	7.4	<LOD	7.4	<LOD-21.6	8.6
Deoxynivalenol	44.3	<LOD-234.7	102.5	<LOD-334.0	97.3	5.4-86.5	92.5	11.7-176.8	92.7	<LOD-261.9	85.9
DON-3-Glucoside	2.9	<LOD-10.2	5.6	<LOD-27.4	5.4	2.34-5.6	5.2	0.6-8.7	5.3	<LOD-11.6	4.9
Zearalenone	5.1	0.2-22.1	4.8	<LOD-26.3	4.5	0.3-6.0	4.1	0.5-7.0	4.0	0.2-7.9	4.5
Nivalenol	15.1	<LOD-65.2	8.0	<LOD-14.0	7.5	<LOD-4.7	6.2	<LOD-5.5	6.0	<LOD-7.9	8.5
Average	172.2		95.6		104.9		98.6		87.9		

**KEYS:** DS- Derived Savannah, HF- Humid Forest, GS-Guinea Savannah, SZ-Savannah Zone and MSZ- Mangrove Savannah Zone.

The result from table 1 shows that the highest incidence of fish feed contamination was recorded for Fumonisin B<sub>1</sub> with mean of 900.9µg/kg (range of <LOD- 6097.0 µg/kg), followed by Fumonisin B<sub>2</sub> of mean 220.6 µg/kg (range of < LOD- 1203.3 µg/kg) and then Aflatoxin B<sub>1</sub> of mean 103.0 µg/kg (range <LOD-418.7 µg/kg), while Zearalenone of mean 4.5 µg/kg ( range <LOD-26.3 µg/kg) had the least. These values are above the European Union permissible values. The co-occurrence of Fumonisin and Zearalenone in the fish feed samples agrees with the findings of Pietsch *et al.*, (2013) who reported that both mycotoxins were firstly reported in samples of commercial fish feeds sampled from Central Europe. While Hashimoto *et al.* (2003) earlier reported the presence of both aflatoxins and fumonisins in 23.8% of the 42 fish feed samples. These are evidences that fish feeds are carriers of multi-mycotoxins as they are majorly formulated from plant sources that are good substrates for moulds proliferation and subsequent mycotoxins production.

**Table 2: T-value, mean and Standard deviation of concentration of multi-mycotoxins in the fish feed samples**

MYCOTOXINS	MEAN ± STD ERROR	STD DEV	T-VALUE
Aflatoxin B <sub>1</sub>	1.03 ± 11.80	116.22	8.76**
Aflatoxin B <sub>2</sub>	12.52 ±1.49	14.72	8.38**
Aflatoxin G <sub>1</sub>	35.99±4.74	8.71	7.60**
Aflatoxin M <sub>1</sub>	2.80±0.88	8.02	5.55**
Fumonisin B <sub>1</sub>	8.59±124.37	1224.90	6.91**
Fumonisin B <sub>2</sub>	2.00±33.46	329.51	5.96**
Fumonisin B <sub>3</sub>	34.00±5.21	51.31	6.52**
HFB <sub>1</sub>	5.56±1.00	9.74	5.62**
DON	67.29±7.26	71.53	9.26**
DON3GluCOSIDE	4.63±1.02	10.07	4.52**
Zearalenone	5.58±1.15	11.31	4.86**
Nivalenol	5.33±1.20	11.81	4.45**

Results from table 2 shows Standard deviation, Standard error and Mean difference of the multi-mycotoxin contaminations in fish feed samples from different Agro-Ecological Zones (AEZs) in Nigeria containing Fumonisins, Aflatoxins, Deoxynivalenol, Zearalenone etc whose presence are of concern in the feed industry. The highest mean value recorded in table 2 was 67.29±7.26 from DON however, highest standard error and standard deviation was from Fumonisin B<sub>1</sub>. Fumonisin B<sub>1</sub> also had the highest toxin value with the range of 0.800-6097µg/kg from Guinea Savannah AEZ of Nigeria. This is in accordance with the observation of Barbosa *et al.*, (2013) who reported that high percentage of the samples (98%) was contaminated with FB<sub>1</sub> at high levels. The following mycotoxins were present in all the fish feed samples; Enniatin B, Equisetin, Beauverucin, Emodin, Alternaric methylether, Methyl sterigmatocystin and Averufin toxins while Ochratoxin, T-2 and HT-2 toxins were not detected in any of the sample.

The highly significant effects of the multi-mycotoxin indicated variation in geographical and ecological distribution of the toxigenic moulds and the mycotoxins produced. This is in accordance with the findings of Cardwell and Cotty, (2002) and Atehnkeng *et al.*(2008).

In this study, the co-occurrence of multi-mycotoxins of concern in food and feed safety calls for urgent check and policies to standardize the quality of fish feeds utilized in fish farming industry.

## REFERENCES

- Abdelhamid AM, Khalil FF, Ragab MA (1998) Problem of mycotoxins in fish production. *Egyptian J Nutr Feeds* 1:63–71
- Ajani, K. 2008. “Sustainable Fish Production, Processing and Marketing.” In *Food Production and Food Security—The Role of Local Government Councils and the Rural Farmers in Agriculture, Food Production and Food Security, July 2008, 140-189pp.*
- Ali N, Sardjono, Yamashita A, Yoshizawa T. (1998). Natural co-occurrence of aflatoxins and Fusarium mycotoxins (Fumonisin, Deoxynivalenol, Nivalenol and Zearalenone) in corn from Indonesia. *Food Addit Contam Part A*15(4):377–384.
- Banu, N. and Muthumary, J. (2008). Screening of commercial feed samples for the presence of multimycotoxins. *Indian Journal of Science and Technology*1 (4):
- Cardwell, K. F. and Cotty, P. J. (2002). Distribution of *Aspergillus Section flavi* among field soils from the four Agro ecological zones of the Republic of Benin, West Africa. *Plant Diseases* 86(4), 434-439.
- Constanze Pietsch, Susanne Kersten, Patricia Burkhardt-Holm, Hana Valenta and Sven Dänicke (2013). Occurrence of Deoxynivalenol and Zearalenone in Commercial Fish Feed: An Initial Study. *Toxins*, 5:184-192
- Delgado C.L., Wada N., Rosegrant M.W., Meijer S., Ahmed M. (2003 ) Fish to 2020: supply and demand in a changing world. *IFPRI*; Washington, DC: 2003.
- Falaye, A.E. 2013. Fish: A Jewel in Land and Sea Environment. An inaugural lecture. University of Ibadan
- FDF, 2008. Fisheries statistics of Nigeria—Federal Department of Fisheries Publication.
- Hashimoto, E.H., M.A.do. Santos, E.Y.S. Ono, C. Hayashi, A.P.F.R.L. Bracarense and E.Y. Hirooka (2003). Bromatology and fumonisin and aflatoxin contamination in aquaculture feed of the region of Londrina, State of Parana, Brazil. *Semina: Ciencias Agrarias , Universidade Estadual de Londrina, Brazil* 24 (1) : 123 – 132.
- Joseph Atehnkeng, Peter S. Ojiambo, Matthias Donner, T. Ikotun, Richard A. Sikora, Peter J. Cotty and Ranajit Bandyopadhyay(2008). Distribution and toxigenicity of *Aspergillus* species isolated from maize kernels from three agro-ecological zones in Nigeria. *International Journal of Food Microbiology* 122 : 74.
- Kathik, P.S., Douressamy, K., Ramaraju, K., Devrajan and Sivakumar, C (2009). Modelling the occurrence of *Prostephanus truncates* (Horn) ( *Coleoptera: Botrichidae*) in Malawi. *Resistant Pest Management Newsletter*, Vol 19, No 1. Pp273-283.
- Lumlertdacha S, Lovell RT (1995) Fumonisin-contaminated dietary corn reduced survival and antibody production by channel catfish challenged with *Edwardsiella ictaluri*. *J Aquatic Anim Health*. 7:1–8
- Sulyok,M., Berthiller,F., Krska,R., and Schuhmacher,R. (2006). Development and validation of a liquid chromatography/tandem mass spectrometric method for the determination of 39 mycotoxins in wheat and maize. *Rapid Communications in Mass Spectrometry* 20: 2649-2659.
- Sulyok,M., Krska,R., and Schuhmacher,R. (2007). A liquid chromatography/tandem mass spectrometric multi-mycotoxin method for the quantification of 87 analytes and its application to semi-quantitative screening of moldy food samples. *Analytical and Bioanalytical Chemistry* 389: 1505-1523.
- Tatiana S Barbosa, Carina M Pereyra, Carla A Soleiro, Erica O Dias, Aguida A Oliveira, Kelly M Keller, Pedro PO Silva, Lilia R Cavaglieri and Carlos AR Rosa (2013). Mycobiota and mycotoxins present in finished fish feeds from farms in the Rio de Janeiro State, Brazil. *International Aquatic Research*, pp 5:3.